

TEST SPECIFIC CHECKLIST

Prepared: March 2009

pH Stabilization Procedure for Testing Acute Lethality of Effluent to Rainbow Trout

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Parameter	Specification	Met Specifics?		
		Y	N	NA
General	pH stabilization techniques are add-on procedures used in conjunction with EPS 1/RM/13 on samples of wastewater effluent (Must)
Conditions for Use . . .	All three of the following conditions are met before any pH stabilization procedures are used (Must) :
	1) Concentration of total ammonia is measured on wastewater effluent sample and used in the calculation of un-ionized ammonia at the initial pH (pHi) of the effluent at 15°C (Must)
	2) Wastewater effluent sample previously collected from the same source failed (i.e., > 50% mortality) the rainbow trout acute lethality test (EPS 1/RM/13) (Must)
	3) Un-ionized ammonia concentration in 100% wastewater is <1.25 mg/L at 15°C, or total ammonia concentration is < maximum total ammonia concentration (y) in mg/L determined using the following formula at the initial pH of the wastewater effluent sample at 15°C: $y = 1.25 (10^{(9.564136638 - \text{pH})} + 1) \text{(Must)}$
pH stabilization methods	One of three techniques for pH stabilization is used to control the pH of the sample at the level measured at test initiation (pH i): (1) CO ₂ Injection, (2) Recycling, and (3) pH Controller (Must)
Total Ammonia	Measured (in mg/L) on all wastewater effluent samples submitted for toxicity testing using EPS 1/RM/13 (Must)
Un-ionized Ammonia	Given that "total ammonia" = NH ₃ + NH ₄ ⁺ , un-ionized ammonia is calculated using the following formula (Must) : un-ionized ammonia = (total ammonia) x [1/(1 + 10 ^{pK - pH})] where: - pK = 9.56 at 15°C - pH is the initial pH (pHi) of the wastewater effluent at 15°C - total ammonia is in mg/L as measured for Condition #1, described above
Sample Preparation	All solutions prepared before aeration started (Must) Stabilization of pH starts when pre-aeration initiated (Must)
Pre-aeration (as per EPS 1/RM/13)	Upon preparation, all test solutions and controls for 30 min at a rate of 6.5 ± 1 mL/min-L (Must) Second period if D.O.(measured after initial 30 min. pre-aeration) in highest test concentration (normally 100% effluent) is < 70% or >100%; pre-aeration of all solutions including controls is continued at 6.5 ± 1 mL/min-L until D.O. is 70 - 100% or 90 min., whichever is shorter (Must) Fish randomly placed in test solutions and test initiated after pre-aeration regardless of whether 70 - 100% aeration achieved (Must)
Air Delivery	Clean airstones used for delivery of CO ₂ mix for CO ₂ Injection technique and for delivery of laboratory air in Recycling and pH Controller techniques (Must) Glass pipette used for delivery of CO ₂ gas in pH Controller technique
Test Conditions				
Aeration	Oil-free compressed laboratory air at a controlled rate of 6.5 ± 1 mL/min-L throughout test period (Must)
Vessel Size & Type	Glass aquaria or non-toxic containers; glass aquaria recommended for Recycling technique
pH	pH of each effluent concentration (i.e., 100, 50, 25...) is maintained at the pH value measure at test initiation (before any aeration is started) in each individual exposure concentration and the control (Must)
Results	LC50 not calculated if there is a non-dose related response that may be due to a gradient of pH values observed during testing across concentrations; 100% wastewater effluent sample still acceptable if other validity criteria are met

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<p><u>CO₂ Injection Technique</u> (continued)</p> <p>% CO₂ for pH Stabilization</p> <p>Flow rates</p> <p>pH Measurements . . .</p> <p>Operating Check</p>	<p>Initial % CO₂ mixture required to stabilize pH is based on the measured pH_i (of each test concentration for a multi-concentration test) and alkalinity of 100% test solution (Must)</p> <p>CO₂ Calibration Table (Table 1 in EPS 1/RM/50) is used to estimate the % CO₂ that is applied for a given pH_i and alkalinity, to provide pH control</p> <p>% CO₂ is adjusted by 0.5% increments if there is an upward or downward trend in pH after initiation of aeration with the CO₂ mixture (pH measured within the first 30 minutes of aeration); CO₂ increased if there is an upward trend in pH, and decreased if there is a downward trend</p> <p>Total aeration rate (CO₂ and lab air) is 6.5 ± 1 mL/min · L throughout test in all exposures and control (Must)</p> <p>Adjusted using adjustable valves on flow meters, based on test solution volume and require final % CO₂</p> <p>Flow rates for CO₂ and lab air determined using following equations:</p> <p>(1) combined flow to test vessel (mL/min) = 6.5 mL/min · L x test volume (L)</p> <p>(2) flow rate of CO₂ mix = $\frac{\text{required } \% \text{ CO}_2}{\% \text{ CO}_2 \text{ in mix (15\%)}}$ x combined flow to vessel (1)</p> <p>(3) flow of lab air = combined flow to vessel (1) - flow rate of CO₂ mix (2)</p> <p>Frequent pH measurements and adjustments to flow of CO₂ mix are made during first three hours of the test to stabilize pH (Must)</p> <p>pH measured and recorded immediately before any aeration (pH_i), at t = 0 h (test start, when fish are introduced), and at t = 0.5, 1, 2, 3, 24, 48, 72, and 96 h in all exposure concentrations and control (Must)</p> <p>pH also measured and recorded with any adjustment to CO₂ flow and a subsequent pH reading taken within 30 minutes after the adjustment (Must)</p> <p>Final pH recorded if 100% mortality in a test concentration before test end (Must)</p> <p>Air line tubing is inspected at least once daily to ensure continual delivery of CO₂ mixture and lab air to all test solutions (Must)</p>
<p><u>Recycling Technique</u></p> <p>pH Control</p> <p>Setup for Recycling Technique</p> <p>Controlling pH Drift . . .</p>	<p>Upward drift of pH is controlled by recycling CO₂ in a closed system (i.e., test vessels sealed with lids, and air, containing CO₂, re-circulated in headspace, preventing loss of CO₂ and maintaining pH) (Must)</p> <p>Recycle technique may accentuate decline in dissolved oxygen for samples with high BOD</p> <p>Recycling apparatus is set up as described in Section 2.3.1 and Figures 6 and 7 of EPS 1/RM/50 (i.e., Closed system with specially fabricated "Recycle" lid; aeration pump that generates 6.5 ± 1 mL/min · L; catch flask to prevent condensate from entering flow meter; flow meter; Tygon® tubing; airstone; and siphoning tube for removing samples for observation and physicochemical measurements)</p> <p>Test containers filled to very top with sample to reduce headspace</p> <p>Recycle lid is secured and tightly sealed to the top of the test container by fastening all O-rings and elastics</p> <p>pH stabilization begins in tanks sealed with Recycling lid, during pre-aeration when pump is started and flow meter adjusted</p> <p>After pre-aeration, fish are added and system is re-sealed</p>

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Recycling Technique (continued..) Flow rates pH Measurements ... Operating Check	Aeration rate is 6.5 ± 1 mL/min · L throughout test in all exposures and control (Must) pH measured and recorded immediately before any aeration (pH i), at t = 0 h (test start, when fish are introduced), and at t = 24, 48, 72, and 96 h in all exposure concentrations and control (Must) ; pH measurements at t = 0.5, 1, and 2 h are also recommended pH also measured and recorded any time the test container is opened (Must) Final pH recorded if 100% mortality in a test concentration before test end (Must) Care is taken when using the siphoning tube for sampling to avoid loss of sample (Must) Dead fish are removed every 24 h, quickly to prevent pH drift (Must) Visual checks are made at least once daily to ensure air lines, pumps, and flow meters are working properly (Must)			
pH Controller Technique pH Control Setup for pH Controller Technique pH Controller Controlling pH Drift ... Flow rates	Upward drift of pH is controlled by aerating wastewater test solutions (including control) using pure CO ₂ (or a mixture of 15% CO ₂ , 21% O ₂ , and 64% N ₂ with separate lines for lab air addition; CO ₂ addition is regulated by a controller that is triggered by a drift in pH above a programmed set point (Must) Apparatus for Controller Technique is set up as described in Section 2.4.1 and Figures 8 to 13 of EPS 1/RM/50 (i.e., CO ₂ is delivered to test vessels from a compressed gas cylinder containing CO ₂ , via gas cylinder regulators, and individual pressure regulators with needle valve assemblies, connected to the gauge assembly (manifold); solenoids, one for each exposure concentration, are used to control the flow of CO ₂ ; and pH controllers, one for each exposure concentration, to monitor and regulate CO ₂ delivery through backflow valves and glass pipettes) Oil or grease is not used on any regulator or cylinder fittings (Must) All solenoids are turned off before the valve on the CO ₂ cylinder is opened (Must) Valve on CO ₂ cylinder is opened and pressure adjusted to 40 psi Working pressure on solenoid is adjusted to 20 psi (i.e., solenoid regulator gauge reads 20 psi) Connections are tested for leaks pH Controller is calibrated daily using certified pH standards (Must) Sensitivity of pH Controller is set before test initiation (± 0.2 pH units) (Must) .. CO ₂ tubing is removed from the exposure solution during calibration (Must) .. Meter calibration is completed rapidly to prevent pH drift (Must) Instructions for calibration and maintenance are provided by manufacturer and reviewed before test initiation One pH probe and controller is used for each test solution for test duration ... Probe is secured 3-5 cm below the surface of the test solution CO ₂ delivery pipette is directly beneath the pH probe (for accurate pH control) Back-flow prevented using back-flow check valve Durable pH probes used to reduce risk of KCl leaks pH stabilization begins during pre-aeration when CO ₂ cylinder is opened Aeration rate for delivery of lab air through airstone is 6.5 ± 1 mL/min · L throughout test in all exposures and control (addition of CO ₂ will slightly increase aeration rate when pH Controller cycles on) (Must)			

